Comparative binding properties of linear and cyclic δ-selective enkephalin analogues: [³H]-[D-Thr², Leu⁵] enkephalyl-Thr⁶ and [³H]-[D-Pen², D-Pen⁵] enkephalin

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The range of δ -selectivity of linear and cyclic analogues of enkephalin in rat brain was found to be: $[D-Pen^2, L-Pen^3]$ enkephalin (DPLPE) > $[D-Pen^2, D-Pen^3]$ enkephalin (DPDPE) > $[D-Thr^2, Leu^3]$ enkephalyl-Thr⁶ (DTLET) > $[D-Ser^2, Leu^3]$ enkephalyl-Thr⁶ (DSLET). Saturation experiments performed with $[^3H]DPDPE$ and $[^3H]DTLET$ in NG108-15 cells and rat brain showed similar binding capacities for both the ligands, but the δ -affinity of $[^3H]DTLET$ ($K_D \approx 1.2$ nM) was much better than that of $[^3H]DPDPE$ ($K_D \approx 7.2$ nM). The rather low δ -affinity of DPDPE induced high experimental errors cancelling the benefit of its better δ -selectivity. Binding experiments in rat or guinea-pig brains showed, in both cases, the better δ -selectivity of $[^3H]DTLET$ compared to $[^3H]DSLET$. The former peptide remains at this time the most appropriate radioactive probe for binding studies of δ -receptor.

 δ -Enkephalin analogue Opioid receptor binding Rat brain Guinea-pig brain NG108-15 hybrid cell

1. INTRODUCTION

Considerable interest has been devoted to the biochemical and pharmacological properties of the δ opioid receptor type [1]. However, the lack of fully δ -selective ligands has led to controversy [2] about its molecular nature, i.e., whether it is an independent receptor [3,4] or μ -interconvertible conformation [5] or allosteric site [6,7]. From both extensive conformational studies performed by NMR and theoretical calculations, Fournié-Zaluski et al. [8] proposed structural requirements for ligands to preferentially interact with μ or δ sites. This led to

Abbreviations: DAGO, [D-Ala²,(Me)Phe⁴,Gly-ol⁵]-enkephalin; DADLE, [D-Ala²,D-Leu⁵]enkephalin; DSLET, [D-Ser²,Leu⁵]enkephalyl-Thr⁶; DTLET, [D-Thr²,Leu⁵]enkephalyl-Thr⁶; DPDPE, [D-Pen²,D-Pen⁵]-enkephalin; DPLPE, [D-Pen²,L-Pen⁵]enkephalin

the syntheses Tyr-D-Ser-Gly-Phe-Leu-Thr (DSLET) and Tyr-D-Thr-Gly-Phe-Leu-Thr (DTLET), which have also been in tritiated form [9,10]. Owing to their enhanced δ -selectivity, these ligands have therefore been used, in preference to the poorly selective [D-Ala²,D-Leu⁵]enkephalin [10], to elucidate the pharmacological role of the δ receptor [11-15], its distribution in the rat brain [16] and its binding properties [17]. Recently, Mosberg et al. [18] described two conformationally restricted cyclic enkephalin analogues, [D-Pen², D-Pen⁵]enkephalin (DPDPE) [D-Pen²,L-Pen⁵]enkephalin (DPLPE), which show an even better selectivity for the δ -binding site than the linear peptides DSLET and DTLET. All of these analogues exhibit very low potencies at xsites [22]. As one of the cyclic compounds, [3H]DPDPE, is now commercially available, it was of interest to study its binding properties on rat

brain tissue which contains both μ and δ receptor types, with the aim of performing competition experiments with selective μ -agonists.

2. MATERIALS AND METHODS

2.1. Chemicals

DSLET [19] and DTLET [10] were prepared as previously described. DPDPE and DPLPE were a generous gift from Dr H.I. Mosberg or purchased from Bachem, AG. [³H]DTLET (45 Ci/mmol) was obtained from its 2,5-dibromotyrosyl precursor [20]. [³H]DSLET (30.5 Ci/mmol) was purchased from New England Nuclear and [³H]DAGO (60 Ci/mmol) and [³H]DPDPE (35 Ci/mmol) from Amersham.

2.2. Preparation of crude membrane fractions and binding assays

The experiments were performed on male Sprague-Dawley rats and guinea-pigs (Ruvel, France). NG108-15 cells were cultivated in Dulbecco's modified Eagle's medium. Crude membrane preparations and binding assays were previously described [17]. NG108-15 cells were homogenized with a Polytron grinder, prior to centrifugation at $100\,000 \times g$ for 35 min. All binding assays were carried out in triplicate in a volume of 1 ml, at 35° C for 45 min unless otherwise indicated.

The $K_{\rm I}$ values were calculated from the relationship $K_{\rm I} = {\rm IC}_{50}/(1 + L/K_{\rm D})$ where L is the concen-

tration of the labelled ligand and K_D its equilibrium dissociation constant. Linear regression analyses were done on a Hewlett-Packard HP85A desk calculator.

3. RESULTS

3.1. Selectivities of bis-penicillamine enkephalins Selectivities of linear and cyclic enkephalin analogues for μ and δ sites of rat brain were indirectly determined by inhibition experiments using [³H]DAGO (1 nM), a μ -selective ligand [21] and [³HIDSI ET (2 nM), a pearly δ -selective ligand

ing [3 H]DAGO (1 nM), a μ -selective ligand [21] and [3 H]DSLET (2 nM), a nearly δ -selective ligand [9,10] (table 1). The Hill coefficients accounted for the homogeneity of the displacement curves observed.

As shown in table 1, bis-penicillamine enkephalins DPDPE and DPLPE had higher K_I values at the μ -binding site than DSLET and DTLET. However, in agreement with a previous study [18], the conformationally restricted enkephalin analogues showed weaker inhibition potencies at δ -sites than the more flexible linear peptides. As illustrated by the ratio of their respective K_I values for δ and μ sites, the selectivity of the studied peptides (table 1) increased over the following range: DSLET, DTLET, DPDPE, DPLPE. These findings disagree with other papers [4,18,22] reporting a better selectivity for DSLET than DTLET.

Table 1
Inhibitory potencies of enkephalin analogues on the binding of [3 H]DAGO (1 nM) at μ -site and of [3 H]DSLET (2 nM) at δ -site in rat brain tissue

Analogues	μ-site			$K_1(\delta)$	
	K ₁ ^a (nM)	n Hill ^a	K _I ^a (nM)	n Hill ^a	Κ ₁ (μ)
DPLPE	873 ± 210	0.83 ± 0.09 (4)	10.9 ± 1.2	0.85 ± 0.05 (5)	0.0125
DPDPE	993 ± 151	0.80 ± 0.03 (3)	19.2 ± 1.4	0.82 ± 0.05 (4)	0.0193
DTLET ^b	25.3 ± 2.5	$0.85 \pm 0.03 (5)$	1.35 ± 0.15	$0.86 \pm 0.02 (5)$	0.053
DSLET ^b	31.0 ± 5.0	0.80 ± 0.05 (4)	4.80 ± 0.80	0.81 ± 0.04 (4)	0.15

a Means ± SE

Results were obtained at 35°C from the analysis of Hill plots using 8 concentrations of unlabelled compound. The number of observations is given in parentheses

b Values from [10]

While DPDPE was approx. 2.5-times more δ selective than DTLET, the latter displayed a
14-times higher apparent affinity for the δ receptor
type.

3.2. Binding characteristics of [³H]DPDPE and [³H]DTLET on rat brain and NG108-15 cells membranes

In the case of [3 H]DPDPE (10 nM), steady-state conditions were established from association kinetics on rat brain membranes at 35 and 4°C. Equilibrium was reached in 45 min at 35°C (k_{+1} (app) = 0.061 min⁻¹) and in 2 h at 4°C (k_{+1} (app) = 0.020 min⁻¹). However, the specific binding at 4°C was only 51% of that at 35°C. The binding kinetics of [3 H]DTLET have already been determined [10].

Saturation experiments were performed on membranes from rat brain and NG108-15 cells, at 35° C. The equilibrium dissociation constants ($K_{\rm D}$) and binding capacities ($B_{\rm max}$) were calculated from linear regression analysis of the saturation isotherms by either Scatchard or Eadie-Hofstee plots. The accuracy in the determination of binding parameters can be evaluated from the standard deviation of the error of the raw data, SD(Erad) [23]. As shown in table 2, in rat brain the SD(Erad) for [3 H]DTLET was around 5% while it reached 20% for [3 H]DPDPE. The range of error for the two radioligands is illustrated in fig.1 by the dif-

ferences in B_{max} and K_{D} values derived from least-squares linear regression analyses of Scatchard and Eadie-Hofstee plots [24]. Since the specific binding of [${}^{3}\text{H}$]DPDPE was greater in NG108-15 cells, its K_{D} values derived from the two analyses were similar, leading therefore to an accurate SD(Erad) estimate of 5%. As shown in table 2, [${}^{3}\text{H}$]DPDPE had one sixth of [${}^{3}\text{H}$]DTLET affinity. The binding capacity of [${}^{3}\text{H}$]DPDPE in rat brain showed no significant difference from [${}^{3}\text{H}$]DTLET binding capacity.

3.3. Binding of [3H]DSLET and [3H]DTLET in rat and guinea-pig brains

Saturation experiments of [3 H]DSLET and [3 H]DTLET showed that in both preparations [3 H]DTLET exhibited the highest affinity: $K_D = 1.36 \pm 0.12$ nM against 3.49 ± 0.22 nM for [3 H]DSLET in rat brain and $K_D = 1.08 \pm 0.08$ nM against 1.97 ± 0.16 nM for [3 H]DSLET in guineapig brain. In accordance with its higher crossreactivity, the capacity for [3 H]DSLET was 29% greater than for [3 H]DTLET in rat brain (1 H 2 H) and 1 H 2 H 3 H $^$

Table 2

Binding parameters of [3H]DPDPE and [3H]DTLET in crude membrane fractions of rat brain and NG108-15 cells at 35°C

³ H-labelled	Tissue	Scatchard analysis		Eadie-Hofstee analysis		
ligand (nM) ^b		K _D ^a (nM)	B _{max} ^a (fmol/mg)	K _D ^a (nM)	B _{max} ^a (fmol/mg)	SD(Erad) ^a (%)
DPDPE (1-20)	Rat brain NG108-15	$ \begin{array}{r} 10.5 \pm 2.4 \\ 7.28 \pm 0.05 \end{array} $	118 ± 11 419 ± 13	5.68 ± 0.63 7.10 ± 0.14	86 ± 3 410 ± 11	20.0 ± 2.9 (4) 5.2 ± 1.3 (3)
DTLET (0.5-7)	Rat brain NG108-15	1.19 ± 0.14 1.25 ± 0.09	103 ± 5 430 ± 10	1.12 ± 0.12 1.19 ± 0.08	101 ± 4 419 ± 9	5.1 ± 1.2 (4) 4.7 ± 1.1 (3)

a Means ± SE

The number of observations is given in parentheses

^b Concentration range of tritiated ligand used (6 concentrations in a volume assay of 0.5 ml)

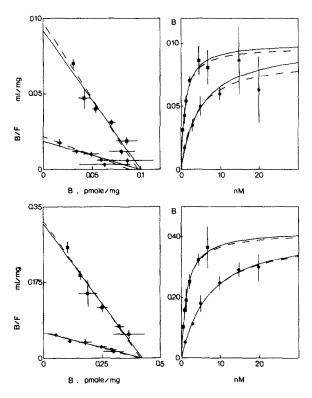


Fig.1. Scatchard (solid lines) and Eadie-Hofstee (dotted lines) analyses of [3H]DTLET (11) and [3H]DPDPE (10) specific binding: results from one experiment performed on the same homogenate of rat brain (upper plots) or NG108-15 cells (lower plots). In the Scatchard representation, B/F is plotted as a function of B(B/F = $B_{\text{max}}/K_{\text{D}}-(1/K_{\text{D}})\cdot \mathbf{B});$ in the Eadie-Hofstee representation, B is plotted as a function of B/F (B = $B_{\text{max}} - K_{\text{D}} \cdot (B/F)$ (inverted axes). Thus, independent variables are, respectively, B and B/F. The corresponding saturation curves are shown on the right side. The concentration ranges were 25-84 and 9-67% of saturation for [3H]DTLET and [3H]DPDPE, respectively.

4. DISCUSSION

Two main molecular approaches for designing δ -probes were successfully used by Fournié-Zaluski et al. [8] and Mosberg et al. [25]. The highly conformationally restricted enkephalin analogues DPDPE and DPLPE exhibit an enhanced δ -selectivity but unfortunately this is associated with a large decrease in affinity. This could indicate that the loss of flexibility may not be favourable for the transconformational binding process [26]. The low

specific binding of [3 H]DPDPE, 26% at 67% saturation, led to large standard deviations compared to [3 H]DTLET which displayed 79% specific binding at 63% saturation. The use of low temperature (4°C) does not improve the specific binding of the cyclic peptide. These features probably account for the large difference observed between the K_D and K_I values for DPDPE while these parameters were almost identical for DTLET. Therefore, [3 H]DPDPE cannot be safely used, either for competition studies or for acute determinations of the concentration of δ sites in brain following biochemical lesions or pharmacological manipulations.

In conclusion, [³H]DTLET seems to be the most appropriate radioligand for binding studies and the cyclic analogue should be reserved for pharmacological experiments.

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